N,O‑Nucleosides from Ene Reactions of Nitrosocarbonyl Intermediates with the 3‑Methyl-2-buten-1-ol

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S Supporting Information

ABSTRACT: Nitrosocarbonyl intermediates undergo ene reactions with allylic alcohols, affording regioisomeric adducts in fair yields. Nitrosocarbonyl benzene reacts with 3-methyl-2-buten-1-ol and follows a Markovnikov orientation and abstracts preferentially the twix hydrogens over the lone ones. With the more sterically demanding nitrosocarbonyl mesitylene and anthracene, the Markovnikov directing effect is relieved and lone abstraction is observed, affording the 5-hydroxy-isoxazolidines that serve as synthons for the preparation of N,O-nucleoside analogues according to the Vorbrüggen protocol.

■ INTRODUCTION

The discovery of several new series of nucleosides analogues with antiviral activity altered the classical way of thinking about nucleosides analogues as antiviral agents.¹ These derivatives play a fundamental role in viral chemotherapy and modification of the sugar fragment and its replacemen[t](#page-9-0) with a carbocyclic moiety resulted in the syntheses of interesting nucleosides analogues that have shown remarkable activity toward a variety of viruses.²

Methods for the synthesis of sugar- and sugar-modified and carbocycli[c-](#page-9-0)nucleosides were studied extensively.³ However, various synthetic problems were frequently encountered, such as low yields, low stereoselectivity, and toxicity pro[b](#page-9-0)lems of the obtained compounds, which are just some of the problems that characterize these synthetic approaches.

Among the various synthetic strategies, the heteroatom substitution produced a series of compounds where the sugar moiety was replaced by alternative heterocyclic rings. Furthermore, the introduction of a side chain on the sugar led to several branched nucleosides that were found to be potential antitumoral or antiviral agents.⁴ In this field, the Chiacchio and Romeo groups have described the synthesis of 4′-α-C-branched N,O-nucleosides based [o](#page-9-0)n the 1,3-dipolar cycloadditions of nitrones with vinyl acetate, followed by coupling with silylated nucleobases. The obtained compounds were evaluated for their activity against various viruses as well as cytoxicity and apoptotic activity.

Recently, we have proposed the synthesis of a new class of carbocyclic nucleosides starting from cyclopentadiene using the nitrosocarbonyl intermediates $(RCONO, 1)$ chemistry.⁶ These intermediates, generated by the mild oxidation of nitrile oxides with tertiary amine N-oxides or by oxidation of hyd[ro](#page-9-0)xamic acids, are efficiently trapped by cyclopentadiene (Scheme 1) to afford the hetero Diels−Alder (HDA) cycloadducts 2 that are highly reactivity dipolarophiles and were employe[d](#page-1-0) to synthesize the conformationally restricted carbocyclic moiety aminols 4 through amide hydrolysis and N−O bond cleavage of the cycloadducts 3. ⁷ Aminols 4 were useful for the linear construction of purine and pyrimidine nucleosides 5. These nucleosides are cha[ra](#page-9-0)cterized by the presence of a secondary hydroxy group on the cyclopentane moiety and by an isoxazoline ring fused to the carbocyclic unit and were found moderately active against human Herpes (types 1 and 2) and Varicella viruses.⁸ Recently, anthryl derivatives were found active with no cellular toxicity at the dose tested against the Hum[a](#page-9-0)n Papilloma virus (HPV).⁹

Applications of nitrosocarbonyls to the syntheses of biologically active molecules ar[e](#page-9-0) well-known, essentially due, first, to the variety of generating methods available (oxidation of hydroxamic acids¹⁰ and nitrile oxides¹¹ and thermal¹² or photochemical¹³ cycloreversions of 1,2,4-oxadiazole-4-oxides)

Received: Oc[tob](#page-9-0)er 29, 2012 Published: December 17, 2012 Scheme 1. Synthetic Pathway of Isoxazoline-Carbocyclic Nucleosides through Nitrosocarbonyl Chemistry and Linear Construction of the Heterobases

Scheme 2. Ene Reactions of Aromatic Nitrosocarbonyl Intermediates with Trisubstituted Olefins

Scheme 3. Ene Reactions of Aromatic Nitrosocarbonyl Intermediates with the 3-Methyl-2-buten-1-ol and Its Methyl Ether and Acetyl Derivatives

and, second, to the exceptional reactivity of these intermediates in HDA reactions. Nevertheless, nitrocarbonyl intermediates are also powerful enophiles, 14 but no applications of ene reactions toward the synthesis of nucleosides are actually reported.¹⁵ We have detailed [th](#page-9-0)at nitrosocarbonyl intermediates, generated at r.t. by the mild oxidation of nitrile oxides, undergo [cle](#page-9-0)an ene reactions with trisubstituted olefins 9. Allylic hydrogens on the more congested side of the alkene are exclusively abstracted (the "cis effect"), thus resembling the singlet oxygen behavior. Nitrosocarbonyl benzene 8P (P = phenyl) follows a Markovnikov (M) orientation and abstracts preferentially the twix hydrogens over the lone ones. With the more sterically demanding nitrosocarbonyl mesitylene 8M (M

= mesityl), the M-directing effect is relieved and comparable twix and lone abstraction are observed (Scheme 2).¹⁶

On pursuing our research in nitrosocarbonyl ene reactions, we extended the studies to other trisubstituted [ole](#page-1-0)fi[ns](#page-10-0) and, in particular, to allylic alcohols $(R = OH)$. The 3-methyl-2-buten-1-ol was found to be the proper model to investigate the selectivity outcome in the ene reaction with nitrosocarbonyls bearing more sterically demanding substituents. An unexpected isoxazolidine deriving from the anti-Markovnikov (AM) addition to the alcohol is presented as a valuable synthon toward the preparation of new N,O-nucleoside analogues through the Vonbrüggen protocol.

■ RESULTS

Ene Reactions with Nitrosocarbonyl Benzene and Mesitylene. Addition of a dichloromethane (DCM) solution of benzohydroximoyl chloride 6 to a stirred solution of Nmethyl-morpholine N-oxide (NMO, 1.1 equiv) and Et_3N (1 equiv or a catalytic amount) in DCM in the presence of an excess (5 equiv) of 3-methyl-2-buten-1-ol 9d afforded, after standing overnight at r.t., the ene adducts 10Pd and 13Pd (Scheme 3) that were isolated upon chromatographic separation of the reaction mixture in the reported yields (Table 1). [T](#page-1-0)he same procedure was followed for the allylic ether $9e^{17}$ and acetate $9f₁₈$ which were prepared according to the reported syntheses.

Table 1. Yields of Adducts in the Reactions of Nitrosocarbonyls 8P,M with Alkenes 9d−9f

Ar-CONO	alkene	10	11	12	13
8P	9d	50			8
	9e	70			
	9f	60			
8M	9d	50	25		
	9e	63		35	
	9f	56		32	

The structures of all isolated ene adducts rely upon the corresponding analytical and spectroscopic data. In the ¹H NMR spectra, the typical olefinic methylene signals are found in the range between 5.00 and 5.43 δ for adducts 10Pd−10Pf and more shielded between 4.89 and 5.00 δ for the derivative 13Pd. The N-OH group is evident when the spectrum is recorded in DMSO and is found strongly deshielded in the range 8.2−8.4 δ for 10Pd−10Pf and even more (9.3−9.5 δ) for adduct 13Pd. In the reaction with the allylic alcohol 9d, compound 13Pd is related to 10Pd and both derive from the

addition of the nitrosocarbonyl benzene 8P to the lesssubstituted carbon atom of the double bond, in full accordance with a M orientation.^{14,16} Adduct 13Pd derives from 10Pd simply upon water elimination.

When the hydroxy g[ro](#page-9-0)[up](#page-10-0) of the alcohol 9d is protected as an ether in 9e or an ester in 9f, the ene reactions with the nitrosocarbonyl benzene 8P proceed straight to single ene adducts 10Pe and 10Pf, respectively, which are obtained in good yields according to the addition of the nitrosocarbonyl electrophilic nitrogen to the alkene in an M fashion.¹⁵

Moving to the nitrosocarbonyl mesitylene 8M case, the experimental conditions were even simpler an[d](#page-10-0) cleaner. Mesitonitrile oxide 7 was added to a stirred solution of NMO (1.1 equiv) in DCM in the presence of an excess (5 equiv) of 3-methyl-2-buten-1-ol 9d at r.t. After stirring overnight and column chromatographic separation, the ene adduct 10Md was isolated in 50% yield along with an unexpected second compound, 11Md, in only, but remarkable, 25% yield.

Compound 10Md is the M adduct, and its structure is in accordance with the analytical and spectroscopic data. The structure of 11Md was not immediately clear. The ¹H NMR spectrum indicates the presence of one deshielded proton at 5.44 δ coupled with two protons at 2.14 and 2.35 δ , while an OH group gives a singlet at 6.98 δ , as confirmed by the corresponding IR spectrum. The ¹³C NMR spectrum presents a signal at 95.3 δ resembling an acetal-type carbon atom. The structure assignment as shown in Scheme 3 came from the Xray analysis (see the Supporting Information). These types of 5 hydroxy-isoxazolidines are usually prepa[re](#page-1-0)d by reaction of hydroxamic acids and α , β [-unsaturated aldeh](#page-9-0)ydes.²⁰ The origin of the isoxazolidine 11Md was undoubtedly attributed to a relief in the M addition determined by the [mor](#page-10-0)e sterically demanding mesityl group that activates the AM path.¹⁶ As the addition of the nitrosocarbonyl moiety occurs on the more substituted carbon atom of the allylic alcohol 9d, th[e p](#page-10-0)rimary adduct is the enol 14 that evolves into the nonisolable aldehyde 15, which undergoes cyclization to the hemiacetal 11Md (Scheme 4). In fact, when the protected alcohols 9e,f were used and the reactions were carried out in the same conditions, adducts 10Me and 10Mf were obtained in 63% and 56% yields, respectively, along with the enol ethers 12Me and 12Mf in 35% and 32% yields, respectively (Scheme 3 and Table 1).

The structures of the adducts obtained from alkenes 9e,f rely upon their analytical and spectrosco[pic](#page-1-0) data. The ¹H NMR spectra of adducts 10Me and 10Mf show the olefinic methylenes as singlets at 4.90 and 5.00 δ for 10Me and at 4.96 and 5.04 δ for 10Mf. The N-OH singlets are found at 9.07

Scheme 6. Synthesis of Isoxazolidine-Nucleoside Analogues through the Vorbrüggen Protocol

and 9.27 δ for the two products, respectively. The enol-ether 12Me showed the double bond signals at 5.23 and 6.52 δ and the N-OH singlet at 9.10 δ while the enol-acetate 12Mf showed the double bond signals at 5.88 and 7.21 δ and the N-OH singlet at 9.38 δ .

The results of the reactions of nitrosocarbonyls 8P,M and olefins 9d−9f seem to show that hydrogen bonding is ineffective with respect to the selectivity. To verify experimentally the influence of the substitution pattern on the reactivity of the double bond, we performed control experiments by means of intermolecular competition reactions of the alkenes 9d versus 9e and 9f, respectively, in the presence of the nitrosocarbonyl benzene 8P and mesitylene 8M. The results showed that the expected compounds (as shown in Table 1) from the competitive reactions were isolated in all the

cases with the same ratios (see the Experimental Section). The allylic alcohol 9d was not found somehow more reactive than the others, and the selectiviy outc[omes were maintained](#page-5-0).

Enforcing the AM Path. We explored the ene reaction of another typical stable nitrile oxide, 21 the anthracenenitrile oxide 17 obtained from the 9-anthraldeyde oxime 16 according to the preparation reported in the literat[ure](#page-10-0)²² by treating a chloroform solution of 16 with NCS and catalytic pyridine at 0 °C for 2 h. The solid nitrile oxide 17 was add[ed](#page-10-0) portionwise to a stirred DCM solution of 5 equiv of the allylic alcohol 9d in the presence of 2 equiv of NMO at r.t. for 48 h. After the usual workup, the residue was submitted to column chromatography, and two products were isolated and identified as compounds 19 and 20 in 27% and 52% yields, respectively (Scheme 5).

Their structures rely upon the relative spectroscopic data. In particular, the ¹H NMR spectrum of the major compound 20 showed a deshielded proton as a triplet at 5.22 δ , coupled with two protons at 2.20 and 2.44 δ . Just before the aromatic signals (7.5−8.6 δ), a doublet was found at 6.84 δ corresponding to the OH group (exchange with D_2O) and whose presence was confirmed by the IR spectrum $(\nu_{\text{OH}} = 3200 \text{ cm}^{-1})$. The ¹³C NMR spectrum showed significantly the acetal-type carbon at 95.8 δ.

The experiment performed with anthracene nitrosocarbonyl 18 and alcohol 9d confirms the relief of the M control observed in the addition of 8M to enes 9d−9f (Table 1), and the results show the remarkable influence of the nitrosocarbonyl substituent on the selectivities. The appro[ac](#page-2-0)h of the bulkier anthracene group determines a strong steric hindrance that disfavors the M path enforcing the AM selectivity.

Synthesis of N,O-Nucleosides. The 5-hydroxy-isoxazolidines 11Bd and 20 obtained from the ene reaction of nitrosocarbonyl mesitylene 8M and nitrosocarbonyl anthracene 18 with the allylic alcohol 9d were used as synthons to prepare N,O-nucleoside analogues by adapting the Vorbrüggen protocol²³ as well as the Chiacchio's procedures for similar compounds.^{5,24} The acetyl derivatives of $21M$, A were prepared accordi[ng](#page-10-0) to standard procedures (Scheme 6).²⁵ The acetylated compound [w](#page-9-0)[as](#page-10-0) obtained in nearly quantitative yield and fully characterized. In the ¹H NMR spectrum of 2[1M](#page-10-0), the presence of the acetyl group is shown by the presenc[e](#page-3-0) of a singlet at 2.09 δ while the OH signal, previously found at 6.98 δ , disappeared. Similarly, in the ${}^{1}H$ NMR spectrum of 21A, the acetyl group is found as a singlet at 2.15 δ while the OH signal at 6.84 δ is now absent.

The Vorbrüggen protocol can be applied on both previously silylated heterobases or commercial compounds in the presence of silylating agents.^{23,26} In the cases at hand, we report the syntheses conducted on commercial heterobases of the uracil family used in the [prese](#page-10-0)nce of the in situ silylating agent, the bis(trimethylsilyl)acetamide (BSA), and trimethylsilyl trifluoromethanesulfonate (TMSO-Tf) as reaction promoter. 27 The acetylated isoxazolidine 21M,A were added under a nitrogen atmosphere at r.t. to a solution of the selected hetero[bas](#page-10-0)es (2 equiv) and BSA (2 equiv), and the solutions became clear after boiling in DCM for a couple of hours. The mixtures were then ice-cooled at 0 °C, TMSO-Tf was added, and the reactions were refluxed overnight (Scheme 6). The desired compounds 22M,A(a−g) were obtained as white solids by simple crystallization or column chromat[og](#page-3-0)raphic purification.

Nucleoside analogues 22M,A(a−g) were isolated from good to excellent yields (48−99%), and their structures rely upon the analytical and spectroscopic data. All the reactions gave single products belonging to the family of uracil derivatives.

The uracil derivative 22Ma is representative of a general trend in the ¹H NMR spectra of this type of compound, and the doublets ($J = 8$ Hz) at 5.67 and 7.64 δ are clearly related to the uracil double bond while the imide NH is found highly deshielded at 10.10 δ as a singlet. The same features were found in cytosine derivative 22Mg. When, in the uracil ring, the $=$ CH−C proton is replaced by a halogen (b, c, d, e) or by a methyl, as in the case of tymine (f), a single proton corresponding to the N-CH= is observed in the range of 7.43−7.80 δ, while the imide NH singlets remain strongly deshielded in the range of 10.08–10.51 δ .

In ¹H NMR spectra of nucleosides 22A(a–g), the signals corresponding to the anthracene and isoxazolidine moieties are

easily detectable. In the ¹H NMR spectrum of the uracil derivative 22Aa, the doublets ($J = 8$ Hz) at 5.49 and 7.86 δ are clearly related to the uracil double bond while the imide NH is found highly deshielded at 11.22 δ as a singlet. In the $^1\rm H$ NMR spectrum of the cytosine derivative 22Mg, the double bond doublets ($J = 8$ Hz) are found at 5.53 and 7.39 δ . Again, when, in the uracil ring, the $=$ CH−C proton is replaced by a halogen (b, c, d, e) or by a methyl, as in the case of tymine (f) , a single proton corresponding to the N−CH= is observed in the range of 7.29−7.86 δ, while the imide NH singlets remain strongly deshielded in the range of 11.23–11.75 δ .

Nucleoside analogues 22M,A can be submitted for biological evaluation without any further structural modification, either for their antiviral behavior or, in other cases, in order to compare their activities with those of reported carbocyclic and heterocyclic structures.^{5,8,24,28} In particular, in nucleoside analogues 22A, the presence of an anthracene ring adds fluorescent properties t[o t](#page-9-0)[hese](#page-10-0) compounds, a further tool for our future studies. The use of fluorescent moieties in the nucleoside synthesis has attracted the interest of various research groups as marker molecules or in the field of imaging to follow their path inside the cells in order to achieve a better understanding of the "in vivo" mechanisms.²⁹ Fluorescent structures can be considered "molecular labels" ³⁰ to be assembled in specific oligofluorosides, as sensors 31 or indicators of the behavior of the DNA.³² New fluorophores are [co](#page-10-0)nstantly developed as well as their applications.^{[3](#page-10-0)3} Fluorescent polyaromatic groups may [be](#page-10-0) active through their ability to establish π -stacking interactions with themsel[ve](#page-10-0)s³⁴ as well as DNA intercalators; 35 these mechanisms are in action in different cases depending upon the biological tar[ge](#page-10-0)ts.

■ DISCUSSION [AN](#page-10-0)D CONCLUSIONS

We have investigated the ene reactions of aromatic nitrosocarbonyls, generated through the mild oxidative protocol with NMO, with allylic alkoxy olefins. The faster oxidation process of nitrile oxide to nitrosocarbonyl intermediate prevents a possible side reaction, that is, the addition of the allylic alcohol to the electrophilic nitrile oxide. From the reaction mixtures, the presence of adducts between the aromatic nitrile oxides and the 3-methyl-2-buten-1-ol 9d was not observed.

The nitrosocarbonyl benzene 8P adds the 3-methyl-2-buten-1-ol 9d, affording adducts 10Pd and 13, which derive from the preferred M path in ene reactions of trisubstituted olefins.^{13,16} If hydroxy-protected allylic ether 9e or ester 9f are used, the reactions proceed straightforwardly to single ene ad[duc](#page-9-0)[ts](#page-10-0) 10Pe,f in accordance with the prevailing HOMO(olefin)− LUMO(nitrosocarbonyl) interaction, somewhat enforced by the polarization of the C $=$ C double bond induced by the CH₂OR group.³⁶ Furthermore, no hydrogen-bonding effects are found in directing the selectivity in the reactions at hand.

Wh[en](#page-10-0) bulkier substituents replace the phenyl group (Figure 1), as in nitrosocarbonyl intermediates 8M and 18, in the ene reactions with the 3-methyl-2-buten-1-ol 9d, the M path is risen [en](#page-5-0)ergetically because of a steric effect and the AM path is favored, affording a mixture of regioisomeric compounds. We have recently detailed the selectivity outcome in ene reactions of nitrosocarbonyl mesitylene 8M with trisubstituted olefins,¹⁶ and the results show the remarkable influence of the nitrosocarbonyl substituent on the selectivities in ene reactio[ns.](#page-10-0) Model calculations on the reaction of 8P,M with tetramethylethylene (TME) shed light on the factors involved in the

Figure 1. Effect of substituents on ene reaction selectivity.

varying selectivities. In these reactions of trimethylethylenes, the M path is favored in the case of nitrosocarbonyl benzene 8P while steric hindrance in the approach of 8M compensates somewhat its electronic preference, and mixtures of M and AM adducts are formed. This mechanism is at work also in the case of 3-methyl-2-buten-1-ol 9d as well as the corresponding hydroxy-protected derivatives 9e,f (Figure 2) and remarkably influences and enforces the selectivity in the reaction of nitrosocarbonyl anthracene 18 with 3-methyl-2-buten-1-ol 9d.

The steric hindrance produced by the anthracene group reverses the selectivity in the ene reaction, and the corresponding 5-hydroxy-isoxazolidine becomes the major product and a convenient intermediate for nucleoside analogue synthesis.

As we have demonstrated, 16 in the reactions of nitrosocarbonyl 8P with TME, the transition structure (TS) of the addition step of 8P to TM[E](#page-10-0) shows no special hindrance between the addends, whereas in the TS of 8M, the mesityl

group is twisted out of the nitrosocarbonyl plane as usual, causing unfavorable steric crowding between its ortho methyl and the "trans" distal TME methyl, as depicted in Figure 2.

This crowding should slow down the M approach of 8M to a trisubstituted ethene and compensates somewhat the M electronic bias. Figure 3 reports the same features in the case of the allylic alcohol 9d. From the M approach point of view, it seems quite clear that [th](#page-6-0)e OH group is not playing any role in directing the addition of the nitrosocarbonyl intermediates, both in the cases depicted in the TS M and M′, as the top view of the structures shows (Figure 3).

The steric clashes at work in the M case drift the addition to the AM path that is the prelu[de](#page-6-0) to the enol formation and subsequent cyclization to the isoxazolidine structures.

The structures of the 5-hydroxy-isozaxolidine 11 and 20 resemble those obtainable through nitrone additions to vinyl ether derivatives and constitute useful synthons toward the prepartion of N,O-nucleoside analogues. By adapting the Vorbrüggen protocol²³ as well as known procedures for similar compounds, $5,24$ we have prepared a selection of uracil derivatives 22M,A(a[−](#page-10-0)g) from good to excellent yields. These products co[ns](#page-9-0)[tit](#page-10-0)ute a new class of nucleoside analogues with a variety of different heterobases, easily inserted on the isoxazolidine ring. Their potentialities in terms of biological activity are totally unexplored, but we believe in future promising results if we compare the new nucleoside structures with those of known compounds, both hetero- and carbocyclic, recently reported in the literature, which were found to be interesting adenosine receptor agonists. 37 The insertion of a fluorecent chromophore allows for planning future applications as fluorescent markers in different biolo[gic](#page-10-0)al targets.

The uracil derivatives $22M$, $A(a-g)$ were sent to the Southern Research Institute (SRI) of Birmingham (AL, USA) within the Tuberculosis Antimicrobial Acquisition and Coordinating Facility (TAACF) program and tested against the Mycobacterium tuberculosis H37Rv in BACTEC 12B medium using the microplate Alamar Blue Assay.³⁸ The minimum inhibition concentration (MIC μ g/mL) was found to be >6.25 for all the compounds with the %Inh in t[he](#page-10-0) range of 22−34. Further antiviral and antitumoral tests are currently under evaluation.

Figure 2. Side view of the B3LYP/6-31G* TS's for the reactions of tetramethylethylene with nitrosocarbonyl benzene and 2,6-dimethylbenzene.¹⁶

Figure 3. Side views and top views of the M and AM ene additions to the 3-methyl-2-buten-1-ol of the nitrosocarbonyl mesitylene.

EXPERIMENTAL SECTION

All melting points (mp's) are uncorrected. Elemental analyses were done on n elemental analyzer available at the Departmen of Chemistryt. ¹H and ¹³C NMR spectra were recorded on a 300 MHz spectrometer (solvents specified). Chemical shifts are expressed in parts per million (ppm) from internal tetramethylsilane (δ) , and coupling constants (J) are in hertz (Hz): b, broad; s, singlet; bs, broad singlet; d, doublet; t, triplet; m, multiplet. IR spectra (Nujol mulls) were recorded on a spectrophotometer available at the Department, and absorbtions (ν) are in cm⁻¹. Column chromatography and tlc: silica gel H60 and GF_{254} , respectively. Eluants: cyclohexane/ethyl acetate 9:1 to pure ethyl acetate; pure CHCl₃ to CHCl₃/MeOH 9/1 for the nucleoside syntheses. The identification of samples from different experiments was secured by mixed mp's and superimposable IR spectra.

Starting and Reference Materials. 3-Methyl-2-buten-1-ol (99%) 9d was purchased from chemical suppliers.

3-Methyl-2-butenyl methyl ether 9e was prepared from 4-bromo-2 methyl-2-butene and MeONa according to the reported synthesis.¹⁷

1-Acetoxy-3-methyl-2-butene 9f was prepared from the alcohol 9d and acetic anhydride as reported.¹⁸

Benzhydroximoyl chloride was obtained by treatment of benzald[ox](#page-10-0)-ime with sodium hypochlorite.³⁹ [Ad](#page-10-0)dition of a slight excess of Et_3N to a DCM solution of benzhydroximoyl chloride furnished in situ BNO.

Mesitonitrile oxide 7 w[as](#page-10-0) obtained by oxidation of 2,4,6 trimethylbenzaldoxime with bromine. 21

The 9-anthraldeyde oxime 16 has been purchased from chemical suppliers.

General Procedure for the Ene [Re](#page-10-0)actions of Nitrosocarbonyl Benzene 8A with Alkenes 9d−9f. To an ice-cooled DCM (200 mL) solution of alkenes 9d−9f (5 equiv) were added 1.9 g (1.3 equiv) of NMO and 2 mL (1.1 equiv) of Et_3N under stirring. A solution of 2.0 g (12.9 mmol) of benzhydroxymoyl chloride 6 in 100 mL of DCM was added dropwise, and the reaction was left under stirring at r.t. for 24 h. After dilution with an equivalent volume of DCM, the organic phase was washed with water and dried over anhydrous $Na₂SO₄$. After filtration, the solvent was then evaporated and the reaction mixture was separated by column chromatography, affording the ene adducts.

10Pd: (1.42 g, 50%) White crystals from ethanol, mp 157−159 °C. IR: ν_{OH} 3450, $\nu_{\text{C=O}}$ 1715 cm⁻¹. ¹H NMR (DMSO) δ: 1.70 (s, 3H, CH₃); 4.70 (m, 2H, CH₂-O); 5.15 (s, 1H + 1H, CH₂); 5.30 (m, 1H, CH-N); 7.2−7.9 (m, 5H, Ph); 8.21 (s, 1H, OH). 13C NMR (DMSO) δ: 23.8; 66.0; 79.9; 120.4; 132.6; 132.9; 133.1; 133.2; 137.5; 142.7; 169.0. Anal. Calcd for C₁₂H₁₅NO₃ (221.25): C, 65.14; H, 6.83; N, 6.33. Found: C, 65.15; H, 6.83; N, 6.25.

10Pe: (2.12 g, 70%) Yellowish oil. IR: ν_{OH} 3203, $\nu_{\text{C=O}}$ 1626 cm⁻¹. . ¹H NMR (DMSO) δ : 1.75 (s, 3H, CH₃); 3.33 (s, 3H, OCH₃); 3.56 (dd, $J = 10$, 4 Hz, 2H, CH-N); 3.74 (t, $J = 10$ Hz, 2H, CH₂-O); 4.89 and 4.97 (s, 1H + 1H, CH₂); 7.3–7.6 (m, 5H, Ph); 9.50 (s, 1H, OH). ¹³C NMR (DMSO) δ: 21.0; 58.1; 69.2; 113.4; 126.0; 127.8; 128.0; 128.2; 128.5; 129.2; 135.4; 140.9; 169.2. Anal. Calcd for $C_{13}H_{17}NO_3$ (235.27): C, 66.36; H, 7.28; N, 5.95. Found: C, 66.35; H, 7.25; N, 5.94.

10Pf: (1.81 g, 60%) Yellowish oil. IR: ν_{OH} 3206, $\nu_{\text{C=O}}$ 1742, 1612 cm[−]¹ . 1 H NMR (DMSO at 80 °C) δ: 1.80 (s, 3H, CH3); 2.01 (s, 3H, CH₃CO); 4.35 (d, J = 7 Hz, 2H, CH-O); 4.91 and 5.00 (s, 1H + 1H, CH2); 5.03 (m, 1H, CH-N); 7.39 (m, 3H, arom.); 7.57 (m, 2H, arom.); 9.35 (s, 1H, OH). 13C NMR (DMSO at 80 °C) δ: 13.5; 20.4; 57.8; 61.9; 113.5; 126.0; 128.3; 128.7; 129.1; 130.0; 140.6; 165.5; 171.5. Anal. Calcd for C₁₃H₁₇NO₃ (235.27): C, 66.36; H, 7.28; N, 5.95. Found: C, 66.34; H, 7.25; N, 5.94.

13Pd: (0.21 g, 8%) Yellowish oil. IR: ν_{OH} 3393, $\nu_{\text{C=O}}$ 1723 cm⁻¹. . ¹H NMR (CDCl₃) δ : 1.90 (s, 3H, CH₃); 5.22 (m, 2H, CH₂); 5.32 and 5.43 (s, 1H + 1H, CH₂); 7.2–8.0 (m, 5H, Ph). ¹³C NMR (CDCl₃) δ : 19.3; 54.1; 118.6; 128.3; 129.3; 129.6; 133.0; 138.5; 154.9; 166.1. Anal. Calcd for $C_{12}H_{13}NO_2$ (203.23): C, 70.91; H, 6.45; N, 6.89. Found: C, 70.85; H, 6.45; N, 6.85.

General Procedure for the Ene Reactions of Nitrosocarbonyl Mesitylene 8M with Alkenes 9d−9f. To an ice-cooled DCM (200 mL) solution of alkenes 9d−9f (5 equiv) was added 1.8 g (1.3 equiv) of NMO under stirring. A solution of 2.0 g (12.4 mmol) of mesitonitrile oxide 7 in 100 mL of DCM was added dropwise, and the reaction was left under stirring at r.t. for 24 h. After dilution with an equivalent volume of DCM, the organic phase was washed with water and dried over anhydrous $Na₂SO₄$. After filtration, the solvent was

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then evaporated and the reaction mixture was separated by column chromatography, affording the ene adducts.

10Md: (1.63 g, 50%) White crystals from ethanol, mp 162−163 °C. IR: ν_{OH} 3163, $\nu_{\text{C=O}}$ 1618 cm⁻¹. ¹H NMR (DMSO) δ : 1.78 (s, 3H, CH₃); 2.14, 2.18, and 2.21 (s, 9H, 3CH₃-Ar); 3.72 (dd, $J = 7$, 6 Hz, 2H, CH₂-O); 4.68 (m, 1H, CH-N); 4.97 (bs, 1H + 1H, CH₂); 5.06 (t, $J = 7$ Hz, 1H, OH); 6.80 (m, 2H, Ph); 9.04 (s, 1H, OH). ¹³C NMR (DMSO) δ: 18.8; 18.9; 20.6; 21.4; 58.8; 60.5; 113.4; 127.3; 133.3; 133.5; 134.6; 136.5; 141.2; 170.0. Anal. Calcd for C₁₅H₂₁NO₃ (263.33): C, 68.41; H, 8.04; N, 5.32. Found: C, 68.47; H, 8.05; N, 5.25.

11Md: (0.82 g, 25%) White crystals from benzene, mp 157−158 °C. IR: ν_{OH} 3150, $\nu_{\text{C=O}}$ 1612 cm⁻¹. ¹H NMR (DMSO) δ : 1.55 and 1.72 (s, 6H, 2CH₃); 2.13, 2.19, and 2.22 (s, 9H, 3CH₃−Ar); 2.14 and 2.35 (s, 1H + 1H, CH₂); 5.44 (m, 1H, CH); 6.79 (m, 2H, Ph); 6.98 (s, 1H, OH). 13C NMR (DMSO) δ: 18.3; 20.6; 25.5; 26.4; 50.2; 60.6; 95.3; 127.2; 127.5; 132.8; 132.9; 134.9; 136.7; 163.2. Anal. Calcd for $C_{15}H_{21}NO_3$ (263.33): C, 68.41; H, 8.04; N, 5.32. Found: C, 68.47; H, 8.06; N, 5.26.

10Me: (2.17 g, 63%) White crystals from ethanol, mp 127−128 °C. IR: ν_{OH} 2920, $\nu_{\text{C=O}}$ 1603 cm⁻¹. ¹H NMR (DMSO) δ: 1.79 (s, 3H, CH₃); 2.16, 2.17, and 2.21 (s, 9H, 3CH₃-Ar); 3.29 (s, 3H, CH₃-O); 3.70 (m, 2H, CH₂-O); 4.90 and 5.00 (s, 1H + 1H, CH₂); 5.24 (m, 1H, CH-N); 6.81 (m, 2H, Ph); 9.07 (s, 1H, OH). 13C NMR (DMSO) δ: 18.6; 18.8; 20.1; 20.6; 21.3; 56.8; 57.8; 68.9; 113.4; 127.3; 133.3; 133.5; 134.5; 136.5; 140.7; 170.0. Anal. Calcd for C₁₆H₂₃NO₃ (277.35): C, 69.28; H, 8.36; N, 5.05. Found: C, 69.26; H, 8.25; N, 5.08.

12Me: (1.20 g, 35%) White crystals from ethanol, mp 133−135 °C. IR: ν_{OH} 2920, $\nu_{\text{C=O}}$ 1656 cm⁻¹. ¹H NMR (DMSO) δ: 1.51 (s, 6H, CH₃); 2.12 and 2.20 (s, 9H, 3CH₃-Ar); 3.46 (s, 3H, CH₃-O); 5.23 (d, J = 13 Hz, 1H, C-CH); 6.52 (d, J = 13 Hz, 1H, O-CH); 6.80 (m, 2H, Ph); 9.10 (s, 1H, OH). 13C NMR (DMSO) δ: 18.5; 20.6; 26.6; 55.5; 61.0; 108.6; 127.2; 132.5; 133.7; 135.9; 136.4; 146.8; 154.5; 170.8. Anal. Calcd for $C_{16}H_{23}NO_3$ (277.35): C, 69.28; H, 8.36; N, 5.05. Found: C, 69.25; H, 8.37; N, 5.07.

10Mf: (2.12 g, 56%) White crystals from ethanol, mp 103−104 °C. IR: ν_{OH} 3086, $\nu_{\text{C=O}}$ 1747, 1658 cm⁻¹. ¹H NMR (DMSO) δ: 1.80 (s, 3H, CH₃); 2.00 (s, 3H, CH₃CO); 2.18 and 2.22 (s, 9H, 3CH₃-Ar); 4.25 and 4.41 (AB syst, 2H, CH₂-O); 4.96 and 5.04 (s, 1H + 1H, CH₂); 5.30 (dd, J = 10, 4 Hz, 1H, CH-N); 6.82 (m, 2H, Ph); 9.27 (s, 1H, OH). 13C NMR (DMSO) δ: 18.7; 18.8; 20.6; 20.7; 21.2; 56.4; 60.8; 114.0; 127.3; 127.4; 133.3; 133.4; 134.2; 136.7; 140.0; 170.1; 170.2. Anal. Calcd for $C_{17}H_{23}NO_4$ (305.36): C, 66.86; H, 7.59; N, 4.59. Found: C, 66.87; H, 7.58; N, 5.00.

12Mf: (1.21 g, 32%) White crystals from ethanol, mp 108−109 °C. IR: ν_{OH} 3098, $\nu_{\text{C=O}}$ 1754, 1667 cm⁻¹. ¹H NMR (DMSO) δ: 1.52 (s, 6H, CH₃); 2.12 and 2.20 (s, 9H, 3CH₃-Ar); 2.12 (s, 3H, CH₃CO); 5.88 (d, J = 13 Hz, 1H, C-CH); 6.79 (m, 2H, Ph); 7.21 (d, J = 13 Hz, 1H, O-CH); 9.38 (s, 1H, OH). 13C NMR (DMSO) δ: 18.5; 20.5; 20.6; 25.8; 60.4; 120.3; 127.2; 132.7; 134.5; 135.9; 136.2; 168.0; 170.8. Anal. Calcd for C₁₇H₂₃NO₄ (305.36): C, 66.86; H, 7.59; N, 4.59. Found: C, 66.85; H, 7.55; N, 5.00.

Ene Reaction of Anthracenenitrile Oxide 18 with 3-Methyl-2-Buten-1-ol 9d. To an anhydrous DCM (80 mL) solution of 3 methyl-2-buten-1-ol 9d (5 equiv) and NMO (2 equiv) was added 2.0 g (9 mmol) of anthracenenitrile oxide 18 portionwise under stirring at r.t. After 48 h, the organic phase was washed with brine and dried over anhydrous $Na₂SO₄$. After filtration and evaporation of the solvent, the residue was submitted to column chromatography. The two main fractions were identified as the adducts 19 and 20, which were characterized as below:

19: (0.79 g, 27%) Pale yellow crystals from ethanol, mp 179−181 °C. IR: ν_{OH} 3254, $\nu_{\text{C=O}}$ 1603 cm⁻¹. ¹H NMR (DMSO) δ : 1.80 (s, 3H, CH₃); 3.35 (s, 2H, CH₂OH); 4.41 (m, 3H, OH); 4.67 (m, 2H, CH₂); 5.49 (b, 1H, CH); 7.57 (m, 4H, arom.); 8.11 (m, 4H, arom.); 8.59 (s, 1H, arom.); 9.71 (s, 1H, OH). 13C NMR (DMSO) δ: 17.9; 25.5; 61.1; 119.6; 123.6; 125.5; 125.7; 126.0; 126.7; 128.4; 129.1; 131.3; 134.6; 155.6. Anal. Calcd for C₂₀H₁₉NO₃ (321.38): C, 74.75; H, 5.96; N, 4.36. Found: C, 74.69; H, 5.94; N, 4.36.

20: (1.52 g, 52%) Straw yellow crystals from ethanol, mp 229 °C (dec.). IR: ν_{OH} 3200, $\nu_{\text{C=O}}$ 1588 cm⁻¹. ¹H NMR (DMSO) δ : 1.83 (s, 3H, CH₃); 1.97 (s, 3H, CH₃); 2.22 (d, 1H, J = 13 Hz, CH); 2.44 (dd, 1H, J = 13, 5 Hz, CH); 5.23 (b, 1H, CH-OH); 6.84 (b, 1H, OH, exch. with D₂O); 7.56 (m, 4H, arom.); 7.89 (m, 1H, arom.); 7.99 (m, 1H, arom.); 8.11 (m, 2H, arom.); 8.62 (s, 1H, arom.). 13C NMR (DMSO) δ: 25.7; 26.6; 50.0; 61.6; 95.8; 124.8; 125.3; 125.5; 125.6; 126.3; 126.4; 126.6; 126.7; 126.8; 128.2; 128.4; 130.6; 130.8; 131.9; 161.8. Anal. Calcd for $C_{20}H_{19}NO_3$ (321.38): C, 74.75; H, 5.96; N, 4.36. Found: C, 74.76; H, 6.00; N, 4.37.

Synthesis of the N-Mesitoyl-3,3-dimethyl-5-acetoxy-1,2 isoxazolidine 21M. To an ice-cooled anhydrous DCM (50 mL) solution of N-mesitoyl-3,3-dimethyl-5-hydroxy-1,2-isoxazolidine 11Md $(3.60 \text{ g}, 14 \text{ mmol.})$ was added 2.2 equiv of Ac₂O added under stirring along with 0.34 equiv of DMAP and 2.2 equiv of $Et₃N$. The reaction was left under stirring at r.t. for 24 h. After dilution with an equivalent volume of DCM, the organic phase was washed with a saturated solution of NaHCO₃ and dried over anhydrous $Na₂SO₄$. After filtration, the solvent was evaporated and a solid was obtained. The acetyl derivative 21M (3.68 g) was obtained in 86% yield and recrystallized from methanol: mp 105−107 °C. IR: $\nu_{\rm C=0}$ 1760 cm⁻¹.
¹H NMR (CDCL) δ: 1.74 and 1.88 (s. 6H 2CH): 2.09 (s. 3H ¹H NMR (CDCl₃) δ : 1.74 and 1.88 (s, 6H, 2CH₃); 2.09 (s, 3H, CH₃CO); 2.28 (s, 9H, 3CH₃-Ar); 2.50 (m, 2H, CH₂); 6.23 (d, J = 6 Hz, 1H, O-CH-OAc); 6.80 (m, 2H, Ph). ¹³C NMR (CDCl₃) δ: 18.4; 18.6; 20.9; 21.0; 49.8; 60.6; 93.9; 127.7; 127.8; 132.8; 133.5; 138.0; 165.1; 169.1. Anal. Calcd for $C_{17}H_{23}NO_4$ (305.36): C, 66.86; H, 7.59; N, 4.59. Found: C, 66.75; H, 7.56; N, 4.58.

Synthesis of the N-Anthracenoyl-3,3-dimethyl-5-acetoxy-**1,2-isoxazolidine 21A.** To an ice-cooled anhydrous DCM (50 mL) solution of N-anthracenoyl-3,3-dimethyl-5-hydroxy-1,2-isoxazolidine 20 (1.40 g, 4.78 mmol.) was added 2.2 equiv of Ac_2O under stirring along with 0.34 equiv of DMAP and 2.2 equiv of $Et₃N$. The reaction was left under stirring at r.t. for 24 h. After dilution with an equivalent volume of DCM, the organic phase was washed with a saturated solution of NaHCO₃ and dried over anhydrous $Na₂SO₄$. The solvent was then evaporated, and a solid was obtained. The acetyl derivative 21A (1.63 g) was obtained in 94% yield and recrystallized from ethanol: mp 200–201 °C. IR: $ν_{C=0}$ 1748 cm⁻¹. ¹H NMR (CDCl₃) δ: 1.84 and 1.97 (s, 3H + 3H, CH₃); 2.15 (s, 3H, CH₃CO); 2.47 (d, 1H, $J = 13$ Hz, H-CH); 2.64 (dd, 1H, $J = 13$, 5 Hz, H-CH); 6.04 (d, 1H, J = 5 Hz, O-CH-O); 7.51 (m, 4H, arom.); 8.04 (m, 4H, arom.); 8.47 (m, 1H, arom.). ¹³C NMR (CDCl₃) δ : 20.8; 26.0; 27.2; 49.5; 61.5; 93.5; 124.7; 124.9; 125.3; 125.8; 126.4; 127.0; 127.5; 127.8; 128.5; 128.6; 130.7; 130.9; 131.1; 164.0; 168.8. Anal. Calcd for $C_{22}H_{21}NO_4$ (363.40): C, 72.71; H, 5.82; N, 3.85. Found: C, 72.71; H, 5.81; N, 3.89.

General Procedure for the Synthesis of Nucleosides 22 by Coupling of Isoxazolidines 21M,A and Heterobases B(a−g). A solution of 2 equiv of heterobases B and 2 equiv of bis(trimethylsilyl) acetamide (BSA) in anhydrous DCM (50 mL) is refluxed under a nitrogen atmosphere for 15−20 min until it becomes clear and hence cooled to ambient. A solution in DCM (10 mL) of isoxazolidines 21M,A (0.20 g, 0.65 mmol) is added dropwise and cooled to 0 $^{\circ}$ C and additioned of 0.12 mL (1 equiv) of TMSO-Tf. The reaction is refluxed under stirring overnight and finally quenched with a saturated solution of NaHCO₃ at pH = 7. The mixture is diluted with an equivalent volume of DCM and washed with water and finally dried over $Na₂SO₄$. From the residues, nucleosides 22M,A(a−g) are isolated upon crystallization or column chromatographic purification.

22Ma: $(0.22 \text{ g}, 95\%)$ White solid from ethanol, mp 215 °C (dec.). IR: ν_{NH} 3166, $\nu_{\text{C=0}}$ 1690, 1642 cm⁻¹. ¹H NMR (CD₃COCD₃) δ: 1.75 and 1.89 (s, 6H, 2CH₃); 2.22 and 2.25 (s, 9H, 3CH₃-Ar); 2.72 and 2.90 (dd, $J = 14$, 7 Hz, 2H, CH₂); 5.67 (d, $J = 8$ Hz, 1H, CO-CH); 6.40 (t, $J = 7$ Hz, 1H, CH); 6.82 (m, 2H, Ph); 7.64 (d, $J = 8$, 1H, CH-N); 10.10 (bs, 1H, NH). ¹³C NMR (CD₃COCD₃) δ: 19.5; 21.4; 25.4; 26.9; 48.8; 64.3; 83.4; 103.9; 129.0; 134.4; 134.9; 139.1; 140.5; 151.6; 154.5; 163.4; 168.0; 170.0. Anal. Calcd for C₁₉H₂₃N₃O₄ (357.40): C, 63.85; H, 6.48; N, 11.76. Found: C, 63.83; H, 6.45; N, 11.75.

22Mb: $(0.24 \text{ g}, 97\%)$ White crystals from acetone, mp 210 °C (dec.). IR: ν_{NH} 3167, $\nu_{\text{C=0}}$ 1709 cm⁻¹. ¹H NMR (CD₃COCD₃) δ : 1.74 and 1.88 (s, 6H, 2CH₃); 2.24, 2.25, and 2.28 (s, 9H, 3CH₃-Ar); 2.76 and 2.90 (dd, $J = 14, 7$ Hz, 2H, CH₂); 6.35 (dt, $J = 7, 1$ Hz, 1H, CH); 6.84 (m, 2H, Ph); 7.70 (d, J = 7 Hz, 1H, CH-N); 10.50 (bs, 1H, NH). ¹³C NMR (CD₃COCD₃) δ: 19.4; 19.5; 21.5; 25.3; 26.9; 49.0; 64.3; 84.1; 124.3; 124.8; 129.1; 134.4; 135.0; 135.4; 139.2; 140.5; 143.6; 150.2; 157.5; 157.8; 167.9. Anal. Calcd for $C_{19}H_{22}N_3O_4F$ (375.40): C, 60.79; H, 5.91; N, 11.19. Found: C, 60.81; H, 5.91; N, 11.21.

22Mc: (0.24 g, 93%) White crystals from acetone, mp 208−211 °C. IR: ν_{NH} 3167, $\nu_{\text{C=0}}$ 1709 cm⁻¹. ¹H NMR (CD₃COCD₃) δ : 1.73 and 1.87 (s, 6H, 2CH₃); 2.25, 2.26, and 2.31 (s, 9H, 3CH₃-Ar); 2.83 and 2.95 (dd, J = 14, 7 Hz, 2H, CH₂); 6.29 (dt, J = 7, 1 Hz, 1H, CH); 6.85 and 6.89 (1, 1H + 1H, Ph); 7.68 (s, 1H, CH-N); 10.60 (bs, 1H, NH). ¹³C NMR (CD₃COCD₃) δ : 19.4; 19.5; 21.5; 25.1; 49.7; 64.2; 84.5; 109.8; 129.1; 129.2; 134.2; 135.0; 135.3; 137.3; 139.3; 150.6; 159.6; 168.2. Anal. Calcd for C₁₉H₂₂N₃O₄Cl (391.85): C, 58.24; H, 5.66; N, 10.72. Found: C, 58.22; H, 5.69; N, 10.71.

22Md: (0.27 g, 95%) White crystals from methanol, mp 114−120 °C. IR: ν_{NH} 3168, $\nu_{C=0}$ 1700 cm⁻¹. ¹H NMR (CD₃COCD₃) δ : 1.72 and 1.86 (s, 6H, 2CH₃); 2.25 and 2.32 (s, 9H, 3CH₃-Ar); 2.84 and 2.96 (dd, J = 14, 7 Hz, 2H, CH₂); 6.28 (t, J = 6 Hz, 1H, CH); 6.87 (m, 2H, Ph); 7.74 (s, 1H, CH-N); 10.51 (bs, 1H, NH). 13C NMR (CD_3COCD_3) δ: 19.5; 19.6; 21.5; 25.1; 26.9; 49.8; 64.2; 84.6; 97.8; 129.0; 129.3; 134.2; 135.0; 135.3; 139.3; 139.8; 150.8; 159.6; 168.3. Anal. Calcd for C₁₉H₂₂N₃O₄Br (436.30): C, 52.31; H, 5.08; N, 9.63. Found: C, 52.25; H, 5.09; N, 9.55.

22Me: (0.29 g, 91%) White crystals from methanol, mp 106−110 °C. IR: ν_{NH} 3169, $\nu_{\text{C=0}}$ 1690 cm⁻¹. ¹H NMR (CD₃COCD₃) δ : 1.72 and 1.87 (s, 6H, 2CH₃); 2.25 and 2.34 (s, 9H, 3CH₃-Ar); 2.82 and 2.96 (dd, J = 14, 7 Hz, 2H, CH₂); 6.27 (t, J = 6 Hz, 1H, CH); 6.85 (m, 2H, Ph); 7.80 (s, 1H, CH-N); 10.45 (bs, 1H, NH). 13C NMR (CD_3COCD_3) δ: 19.5; 19.8; 21.5; 25.0; 26.9; 49.9; 64.2; 69.9; 84.4; 129.0; 129.4; 134.1; 135.0; 135.3; 139.2; 144.7; 151.2; 160.8; 168.4. Anal. Calcd for C₁₉H₂₂N₃O₄I (483.30): C, 47.22; H, 4.59; N, 8.69. Found: C, 47.15; H, 4.55; N, 8.49.

22Mf: (0.19 g, 78%) White crystals from ethyl acetate, mp 99−104 °C. IR: ν_{NH} 3170, $\nu_{C=0}$ 1700 cm⁻¹. ¹H NMR (CD₃COCD₃) δ : 1.74 and 1.88 (s, 6H, 2CH₃); 1.83 (s, 3H, CH₃-C); 2.22, 2.25, and 2.27 (s, 9H, 3CH₃-Ar); 2.74 and 2.86 (dd, J = 14, 7 Hz, 2H, CH₂); 6.39 (t, J = 7 Hz, 1H, CH); 6.82 (m, 2H, Ph); 7.43 (d, J = 1 Hz, 1H, CH-N); 10.08 (bs, 1H, NH). ¹³C NMR (CD₃COCD₃) δ: 12.8; 19.5; 21.4; 25.3; 26.9; 48.6; 64.3; 83.3; 112.1; 129.0; 134.3; 134.9; 135.5; 135.8; 139.0; 151.6; 164.2; 167.7. Anal. Calcd for $C_{20}H_{25}N_3O_4$ (371.42): C, 64.67; H, 6.78; N, 11.31. Found: C, 64.65; H, 6.75; N, 11.25.

22Mg: (0.11 g, 46%) White crystals from ethyl acetate, mp 107− 110 °C. IR: ν_{NH2} 3360, 3170, $\nu_{\text{C=0}}$ 1640 cm⁻¹. ¹H NMR (CD_3COCD_3) δ: 1.75 and 1.83 (s, 6H, 2CH₃); 2.21, 2.24, and 2.26 $(s, 9H, 3CH₃-Ar)$; 2.60 and 2.83 (dd, J = 14, 7 Hz, 2H, CH₂); 5.88 (d, J = 8 Hz, 1H, CH); 6.43 (t, J = 7 Hz, 1H, CH); 6.81 (m, 2H, Ph); 7.38 $(s, 2H, NH₂)$; 7.56 (d, J = 8 Hz, 1H, CH-N). ¹³C NMR (CD₃COCD₃) δ: 19.5; 21.4; 23.6; 25.6; 27.1; 49.5; 64.0; 84.2; 95.7; 128.8; 128.9; 129.0; 129.5; 134.3; 134.9; 135.5; 139.0; 141.4; 156.1; 167.3. Anal. Calcd for $C_{19}H_{24}N_4O_3$ (356.41): C, 64.02; H, 6.79; N, 15.72. Found: C, 63.90; H, 6.65; N, 15.55.

22Aa: $(0.26 \text{ g}, 97\%)$ White solid from ethanol, mp > 200 °C. IR: ν_{NH} 3167, $\nu_{\text{C=O}}$ 1686 cm⁻¹. ¹H NMR (DMSO) δ: 1.84 (s, 3H, CH₃); 2.03 (s, 3H, CH₃); 2.79 (m, 2H, CH₂); 5.49 (d, 1H, J = 8 Hz, CH-CO); 6.17 (dd, 1H, J = 7, 6 Hz, O-CH-N); 7.59 (m, 5H, arom.); 7.86 $(d, 1H, J = 8 Hz, N-CH); 7.99 (m, 1H, arom.); 8.12 (m, 2H, arom.);$ 8.66 (s, 1H, arom.); 11.22 (s, 1H, NH). 13C NMR (DMSO) δ: 25.1; 25.8; 46.2; 63.6; 82.3; 102.3; 124.3; 124.7; 125.6; 126.3; 126.7; 126.8; 126.9; 127.8; 128.5; 128.6; 130.4; 130.5; 140.0; 150.1; 162.6; 164.4. Anal. Calcd for $C_{24}H_{21}O_4N_3$ (415.43) C, 69.38; H, 5.10; N, 10.12. Found: C, 69.39; H, 5.05; N, 10.13

22Ab: (0.26 g, 93%) White solid from ethanol, mp > 200 °C. IR: ν_{NH} 3168, $\nu_{\text{C=O}}$ 1720 cm⁻¹. ¹H NMR (DMSO) δ: 1.83 (s, 3H, CH₃); 2.04 (s, 3H, CH₃); 2.79 (m, 2H, CH₂); 6.16 (t, 1H, J = 7 Hz, O-CH-N); 7.58 (m, 4H, arom.); 7.81 (d, 1H, J = 7 Hz, N-CH); 7.86 (d, 1H, J = 9 Hz, arom.); 8.04 (m, 1H, arom.); 8.13 (m, 2H, arom.); 8.67 (s, 1H, arom.); 11.75 (s, 1H, NH). 13C NMR (DMSO) δ: 24.9; 25.7; 45.9; 63.8; 82.7; 124.0; 124.3; 124.4; 124.6; 125.7; 126.4; 126.6; 126.8; 126.9; 127.8; 128.5; 129.6; 130.5; 130.6; 138.2; 138.4; 141.5; 148.8; 156.5; 156.8; 164.2. Anal. Calcd for $C_{24}H_{20}O_4N_3F$ (433.42) C, 66.51; H, 4.65; N, 9.70. Found: C, 66.51; H, 4.64; N, 9.71.

22Ac: (0.29 g, 97%) White solid from ethanol, mp 185−188 °C. IR: ν_{NH} 3067, $\nu_{\text{C=O}}$ 1728 cm⁻¹. ¹H NMR (DMSO) δ: 1.83 (s, 3H, CH₃); 2.03 (s, 3H, CH₃); 2.84 (m, 2H, CH₂); 6.12 (1H, t, J = 6 Hz, O-CH-N); 7.58 (m, 4H, arom.); 7.72 (s, 1H, N-CH); 7.86 (d, 1H, J = 8 Hz, arom.); 8.00 (d, 1H, J = 8 Hz, arom.); 8.12 (m, 2H, arom.); 8.67 (s, 1H, arom.); 11.78 (s, 1H, NH). ¹³C NMR (DMSO) δ: 24.8; 25.7; 46.3; 63.7; 83.1; 107.9; 124.3; 125.6; 125.7; 126.4; 126.8; 126.9; 127.9; 128.6; 130.4; 130.5; 130.6; 136.7; 149.2; 158.6; 164.4. Anal. Calcd for $C_{24}H_{20}O_4N_3Cl$ (449.89) C, 64.07; H, 4.48; N, 9.34. Found: C, 64.08; H, 4.49; N, 9.31.

22Ad: (0.31 g, 95%) White solid from ethanol, mp 167–170 °C. IR: ν_{NH} 3058, $\nu_{\text{C=0}}$ 1726 cm⁻¹. ¹H NMR (DMSO) δ : 1.83 (s, 3H, CH₃); 2.03 (s, 3H, CH₃); 2.85 (m, 2H, CH₂); 6.12 (t, 1H, J = 6 Hz, O-CH-N); 7.57 (m, 4H, arom.); 7.81 (s, 1H, N-CH); 7.86 (d, 1H, J = 8 Hz, arom.); 7.97 (d, 1H, J = 8, 4 Hz, arom.); 8.13 (m, 2H, arom.); 8.67 (s, 1H, arom.); 11.73 (s, 1H, NH). ¹³C NMR (DMSO) δ: 24.9; 25.7; 46.1; 63.7; 83.1; 96.5; 124.3; 125.6; 126.4; 126.9; 127.8; 128.6; 130.4; 130.5; 139.2; 149.4; 158.7; 164.4. Anal. Calcd for C24H20O4N3Br (494.33) C, 58.31; H, 4.08; N, 8.50. Found: C, 58.31; H, 4.11; N, 8.51.

22Ae: (0.35 g, 99%) White solid from ethanol, mp 164−166 °C. IR: ν_{NH} 3059, $\nu_{\text{C=O}}$ 1724 cm⁻¹. ¹H NMR (DMSO) δ: 1.83 (s, 3H, CH₃); 2.03 (s, 3H, CH₃); 2.86 (m, 2H, CH₂); 6.13 (t, 1H, J = 6 Hz, O-CH-N); 7.58 (m, 4H, arom.); 7.85 (d, 2H, J = 7 Hz, arom.); 7.86 (s, 1H, N-CH); 7.96 (d, 1H, J = 8 Hz, arom.); 8.13 (m, 2H, arom.); 8.67 (s, 1H, arom.); 11.60 (s, 1H, NH). ¹³C NMR (DMSO) δ: 25.0; 25.7; 45.9; 63.7; 70.6; 82.8; 124.3; 125.6; 126.4; 126.9; 127.0; 127.8; 128.6; 130.4; 130.5; 143.9; 149.8; 160.1; 164.4. Anal. Calcd for $C_{24}H_{20}O_4N_3I$ (541.32) C, 53.25; H, 3.72; N, 7.76. Found: C, 53.22; H, 3.72; N, 7.80.

22Af: (0.24 g, 86%) White solid from ethanol, mp 155−157 °C. IR: ν_{NH} 3058, $\nu_{\text{C=O}}$ 1719 cm⁻¹. ¹H NMR (DMSO) δ: 1.63 (s, 3H, CH₃); 1.84 (s, 3H, CH₃); 2.05 (s, 3H, CH₃); 2.78 (m, 2H, CH₂); 6.19 (t, 1H, $J = 7$ Hz, O-CH-N); 7.29 (d, 1H, $J = 1$ Hz, N-CH); 7.59 (m, 4H, arom.); 7.86 (d, 1H, $J = 9$ Hz); 8.04 (m, 1H, arom.); 8.13 (m, 2H, arom.); 8.66 (s, 1H, arom.); 11.23 (s, 1H, NH). 13C NMR (DMSO) δ: 11.5; 24.4; 25.5; 45.9; 63.3; 81.6; 109.7; 123.9; 124.3; 125.2; 126.0; 126.2; 126.4; 126.5; 127.4; 128.1; 128.2; 130.0; 130.1; 134.4; 149.7; 162.9; 163.8. Anal. Calcd for $C_{25}H_{23}O_4N_3$ (429.46) C, 69.91; H, 5.40; N, 9.79. Found: C, 69.91; H, 5.41; N, 9.75.

22Ag: (0.04 g, 48%) White solid from ethanol, mp 188−191 °C. IR: ν_{NH2} 3362, 3171, $\nu_{\text{C=0}}$ 1664 cm⁻¹. ¹H NMR (DMSO) δ: 1.87 (s, 3H, CH₃); 1.98 (s, 1.98, CH₃); 2.59 e 2.84 (m, 1H + 1H, CH₂); 5.53 (d, 1H, $J = 7$ Hz, CH-CN); 6.22 (t, 1H, $J = 6$ Hz, O-CH-N); 7.18 (d, 2H, NH2); 7.39 (d, 1H, J = 7 Hz, N-CH); 7.58 (m, 4H, arom.); 7.88 (d, 1H, J = 8 Hz, arom.); 8.00 (m, 1H, arom.); 8.13 (m, 2H, arom.); 8.66 (s, 1H, arom). 13C NMR (DMSO) δ: 22.8; 25.1; 26.0; 47.2; 63.4; 82.7; 94.7; 124.3; 124.8; 125.6; 125.7; 126.4; 126.6; 126.8; 126.9; 127.7; 128.5; 128.6; 130.5; 130.6; 140.0; 154.4; 163.8; 165.3. Anal. Calcd for $C_{24}H_{22}O_3N_4$ (414.45) C, 69.55; H, 5.35; N, 13.52. Found: C, 69.51; H, 5.31; N, 13.52.

Competition Experiments. Equimolecular amounts (20 equiv) of 3-methyl-2-buten-1-ol and the corresponding acetate and methyl ether were allowed to react with 1 equiv of either benzonitrile oxide or mesitonitrile oxide in the presence of 1.2 equiv of NMO in anhydrous DCM as solvent. After the workup previously described, the reaction mixtures were submitted to chromatographic separation for the isolation and quantification of the ene obtained from the yields given in Table 1 (see Table 2).

Biological Tests. Primary screen (dose response): determination of a 90% inhibitory concentration (IC90). The initial screen is conducte[d](#page-2-0) against My[co](#page-9-0)bacterium tuberculosis H37Rv (ATCC 27294) in BACTEC 12B medium using the microplate Alamar Blue assay (MABA). Compounds are tested in ten 2-fold dilutions, typically from 100 to 0.19 μ g/mL. The IC90 is defined as the concentration effecting a reduction in fluorescence of 90% relative to controls. This value is determined from the dose−response curve using a curve-fitting Table 2

program. Any IC90 value of \geq 10 μ g/mL is considered "Active" for antitubercular activity. For further information, see: http://www.taacf. org.

X-ray Crystallography. A summary of crystal data, data collection, and structure refinement of compounds 11Md and 21A [is given in](http://www.taacf.org) [Tab](http://www.taacf.org)le S2 of the Supporting Information. The structures are solved by direct methods. Non-hydrogen atoms were refined anisotropically, and hydrogen atoms, located from the difference Fourier synthesis, were refined isotropically.⁴⁰

CCDC 905532 for compound 11Md and CCDC 905533 for compound 21A co[nta](#page-10-0)in the supplementary data. These data can be obtained free of charge at www.ccdc.cam.ac.uk/conts/retrieving.html [or from the Cambridge Crystallography Data Centre 12, Union Road, Cambridge CB2 IEZ, U.K., Fax: +44−[1223/336033; E-mail: deposit@](www.ccdc.cam.ac.uk/conts/retrieving.html) ccdc.cam.ac.uk].⁴¹

■ ASSOCI[AT](#page-10-0)ED CONTENT

6 [Supporti](mailto:deposit@ccdc.cam.ac.uk)ng Information

NMR spectra of new compounds, X-ray crystallographic data, and crystallographic data in CIF format. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The auth[ors declare no competin](mailto:paolo.quadrelli@unipv.it)g financial interest.

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■ REFERENCES

(1) (a) Herdewijn, P. Drug Discovery Today 1997, 2, 235−242. (b) Wang, P.; Hong, J. H.; Cooperwood, J. S.; Chu, C. K. Antiviral Res. 1998, 40, 19−44. (c) Piperno, A.; Chiacchio, M. A.; Iannazzo, D.; Romeo, R. Curr. Med. Chem. 2006, 13, 3675−3695.

(2) (a) De Clercq, E. Biochim. Biophys. Acta 2002, 1587, 258−275. (b) De Clercq, E. Med. Res. Rev. 2002, 22, 531−542. (c) Bricaud, A. D.; Herdewijn, P.; De Clercq, E. Biochem. Pharmacol. 1983, 3583− 3588.

(3) (a) Wu, Q.; Simons, C. Synthesis 2004, 1533−1553 and references therein. (b) Rajappan, V. P.; Yin, X.; Schneller, S. W. Tetrahedron 2002, 58, 9889−9895. (c) Ishikura, M.; Murakami, A.; Katagiri, N. Org. Biomol. Chem. 2003, 1, 452−453. (d) Jeong, L. S.; Yoo, S. J.; Lee, K. M.; Koo, M. J.; Choi, W. J.; Kim, H. O.; Park, J. G.; Lee, S. K.; Chun, M. W. J. Med. Chem. 2003, 46, 201−202. (e) Gouverneur, V.; McCarthy, S. J.; Mineur, C.; Belotti, D.; Dive, G.; Ghosez, L. Tetrahedron 1998, 54, 10537−10554. (f) Kirby, G. W.; Nazeer, M. J. Chem. Soc., Perkin Trans. 1 1993, 1397-1402.

(4) (a) Chang, C. N.; Doong, S. L.; Zhou, J. H.; Beach, J. W.; Jeong, L. S.; Chu, C. K.; Tsai, C. H.; Cheng, Y. C. J. Biol. Chem. 1992, 267, 13938−13942. (b) Grove, K. L.; Guo, X.; Lui, S. H.; Gao, Z.; Chu, C. K.; Cheng, Y. C. Cancer Res. 1995, 55, 3008−3011.

(5) (a) Pistara, V.; Corsaro, A.; Chiacchio, M. A.; Greco, G.; ̀ Quadrelli, P. ARKIVOC 2011, 270−285. (b) Chiacchio, U.; Genovese, F.; Iannazzo, D.; Piperno, A.; Quadrelli, P.; Corsaro, A.; Romeo, R.; Valveri, V.; Mastino, A. Bioorg. Med. Chem. 2004, 12, 3903−3909. (c) Chiacchio, U.; Genovese., F.; Iannazzo, D.; Librando, V.; Merino, P.; Rescifina, A.; Romeo, R.; Propopio, A.; Romeo, G. Tetrahedron 2004, 60, 441−448. (d) Saita, M. G.; Chiacchio, U.; Iannazzo, D.; Merino, P.; Piperno, A.; Previtera, T.; Rescifina, A.; Romeo, G.; Romeo, R. Nucleosides, Nucleotides Nucleic Acids 2003, 22, 739−742. (e) Chiacchio, U.; Corsaro, A.; Iannazzo, D.; Piperno, A.; Pistara, V.; ̀ Rescifina, A.; Romeo, A.; Valveri, V.; Mastino, A.; Romeo, G. J. Med. Chem. 2003, 46, 3696−3702.

(6) (a) Quadrelli, P.; Scrocchi, R.; Caramella, P.; Rescifina, A.; Piperno, A. Tetrahedron 2004, 60, 3643−3651. (b) Quadrelli, P.; Mella, M.; Carosso, S.; Bovio, B.; Caramella, P. Eur. J. Org. Chem. 2007, 6003−6015. (c) Quadrelli, P.; Mella, M.; Assanelli, G.; Piccanello, A. Tetrahedron 2008, 64, 7312−7317. (d) Savion, M.; Memeo, M. G.; Bovio, B.; Grazioso, G.; Legnani, L.; Quadrelli, P. Tetrahedron 2012, 68, 1845−1852.

(7) (a) Quadrelli, P.; Mella, M.; Paganoni, P.; Caramella, P. Eur. J. Org. Chem. 2000, 2613−2620. (b) Quadrelli, P.; Fassardi, V.; Cardarelli, A.; Caramella, P. Eur. J. Org. Chem. 2002, 2058−2065.

(8) For antiviral activity, the manuscript is in preparation. See also: (a) Kitade, Y.; Kojima, H.; Zulfiqur, F.; Kim, H. S.; Wataya, Y. Bioorg. Med. Chem. Lett. 2003, 13, 3963−3965. (b) Ramesh, N. G.; Klunder, A. J. H.; Zwanenburg, B. J. Org. Chem. 1999, 64, 3635−3641. (c) Wu, J.; Schneller, S. W.; Seley, K. L.; DeClerq, E. Heterocycles 1998, 47, 757−763. (d) Siddiqi, S. M.; Raissian, M.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; DeClerq, E. Bioorg. Med. Chem. Lett. 1993, 3, 663−666. (e) Da Silva, A. D.; Coimbra, E. S.; Fourrey, J. L.; Machado, A. S.; Robert-Gero, M. Tetrahedron Lett. 1993, 34, 6745−6748. (f) Koga, M.; Schneller, S. W. Tetrahedron Lett. 1990, 31, 5861−5864.

(9) Moggio, Y.; Legnani, L.; Bovio, B.; Memeo, M. G.; Quadrelli, P. Tetrahedron 2012, 68, 1384−1392.

(10) (a) Kirby, G. W. Chem. Soc. Rev. 1977, 6, 1−24. (b) Sato, T.; Aoyagi, S.; Kibayashi, C. Org. Lett. 2003, 5, 3839−3842. (c) Momiyama, N.; Yamamoto, H. Org. Lett. 2002, 4, 3579−3582. (d) Ware, R. W., Jr.; Day, C. S.; King, S. B. J. Org. Chem. 2002, 67, 6174−6180. (e) Chow, C. P.; Shea, K. J.; Sparks, S. M. Org. Lett. 2002, 4, 2637−2640. (f) Surman, M. D.; Mulvihill, M. J.; Miller, M. J. Tetrahedron Lett. 2002, 43, 1131−1134. (g) Surman, M. D.; Mulvihill, M. J.; Miller, M. J. J. Org. Chem. 2002, 67, 4115−4121. (h) Iwasa, S.; Tajima, K.; Tsushima, S.; Nishiyama, H. Tetrahedron Lett. 2001, 42, 5897−5899. (i) Flower, K. R.; Lightfoot, A. P.; Wan, H.; Whiting, A. J. Chem. Soc., Perkin Trans. 1 2002, 2058−2064. (j) Iwasa, S.; Fakhruddin, A.; Tsukamoto, Y.; Kameyama, M.; Nishiyama, H. Tetrahedron Lett. 2002, 43, 6159−6161. (k) Pulacchini, S.; Sibbons, K. F.; Shastri, K.; Motevalli, M.; Watkinson, M.; Wan, H.; Whiting, A.; Lightfoot, A. P. Dalton Trans. 2003, 2043−2052. (l) Adam, W.; Bottke, N.; Krebs, O.; Saha-Moller, C. R. Eur. J. Org. Chem. 1999, 1963−1965.

(11) (a) Quadrelli, P.; Gamba Invernizzi, A.; Caramella, P. Tetrahedron Lett. 1996, 37, 1909−1996. (b) Quadrelli, P.; Mella, M.; Gamba Invernizzi, A.; Caramella, P. Tetrahedron 1999, 55, 10497− 10510.

(12) Quadrelli, P.; Campari, G.; Mella, M.; Caramella, P. Tetrahedron Lett. 2000, 41, 2019−2022.

(13) Quadrelli, P.; Mella, M.; Caramella, P. Tetrahedron Lett. 1999, 40, 797−800.

(14) Quadrelli, P.; Mella, M.; Caramella, P. Tetrahedron Lett. 1998, 39, 3233−3236.

(15) (a) Bodnar, B. S.; Miller, M. J. Angew. Chem., Int. Ed. 2011, 50, 5630−5647. (b) Miller, M. J. Proceedings of the International Symposium on Advances in Synthetic, Combinatorial, and Medicinal Chemistry, Moscow, Russia, May 5−8, 2004; T14 and P140. (c) Vogt, P. F.; Miller, M. J. Tetrahedron 1998, 54, 1317–1348. (d) Boger, L.; Weinreb, S. M. Hetero Diels-Alder Methodology in Organic Synthesis; Academic Press: San Diego, CA, 1987.

The Journal of Organic Chemistry and the Second Second

(16) (a) Quadrelli, P.; Mella, M.; Piccanello, A.; Romano, S.; Caramella, P. J. Org. Chem. 2007, 72, 1807−1810. (b) Quadrelli, P.; Romano, S.; Piccanello, A.; Caramella, P. J. Org. Chem. 2009, 74, 2301−2310. (c) Adam, W.; Krebs, O. Chem. Rev. 2003, 103, 4131− 4146.

(17) Johnston, B. D.; Czyzewska, E.; Oehlschlager, A. C. J. Org. Chem. 1987, 52, 3693−3697.

(18) Kann, N.; Rein, T.; Akermark, B.; Helquist, P. J. Org. Chem. 1990, 55, 5312−5323.

(19) For recent direct observation of acyl nitroso species, see: (a) Cohen, A. D.; Zeng, B.-B.; King, S. B.; Toscano, J. P. J. Am. Chem. Soc. 2003, 125, 1444−1445. (b) Evans, A. S.; Cohen, A. D.; Gurard-Levin, Z. A.; Kebede, N.; Celius, T. C.; Miceli, A. P.; Toscano, J. P. Can. J. Chem. 2011, 89, 130−138.

(20) (a) Motorina, I. A.; Sviridova, L. A.; Golubeva, G. A.; Bundel, Y. G. Tetrahedron Lett. 1989, 30, 117−120. (b) Motorina, I. A.; Sviridova, L. A.; Golubeva, G. A.; Bundel, Y. G. Khim. Geterotsikl. Soedin. 1990, 976−979.

(21) Grundmann, C.; Dean, J. M. J. Org. Chem. 1965, 30, 2809− 2812.

(22) (a) Brochard, L.; Joseph, B.; Viaud, M.-C.; Rollin, P. Synth. Commun. 1994, 24, 1403−1414. (b) Da Ros, T.; Prato, M.; Lucchini, V. J. Org. Chem. 2000, 65, 4289−4297.

(23) (a) Vorbrü ggen, H.; Krolikiewicz, K.; Bennua-Skalmowski, B. U.S. Patent 5,750,676, 1998. (b) Vorbrü ggen, H.; Krolikiewicz, K.; Bennua, B. *Chem. Ber.* **1981**, 114, 1234−1255. (c) Vorbrüggen, H.; Höfle, G. Chem. Ber. 1981, 114, 1256−1268. (d) Vorbrü ggen, H.; Bennua, B. Chem. Ber. 1981, 114, 1279−1286.

(24) (a) Chiacchio, U.; Corsaro, A.; Iannazzo, D.; Piperno, A.; Pistarà, V.; Procopio, A.; Rescifina, A.; Romeo, G.; Romeo, R.; Siciliano, M. C. R.; Valveri, E. ARKIVOC 2002, 159−167. (b) Gotkowska, J.; Balzarini, J.; Piotrowska, D. G. Tetrahedron Lett. 2012, 53, 7097−7100.

(25) For acetylation of 5-hydroxy-isoxazolidines, see: (a) Xiang, Y.; Gong, Y.; Zhao, K. Tetrahedon Lett. 1996, 37, 4877−4880. (b) Zhang, X.; Qing, F.-L; Yu, Y. J. Org. Chem. 2000, 65, 7075−7082. (c) Zhang, X.; Xia, H.; Dong, X.; Jin, J.; Meng, W.-D; Qing, F.-L. J. Org. Chem. 2003, 68, 9026−9033.

(26) Yoshimura, Y.; Kitano, K.; Yamada, K.; Satoh, H.; Watanabe, M.; Miura, S.; Sakata, S.; Sasaki, T.; Matsuda, A. J. Org. Chem. 1997, 62, 3140−3152.

(27) Caddell, J. M.; Chapman, A. M.; Cooley, B. E.; Downey, B. P.; LeBlanc, M. P.; Jackson, M. M.; O'Connell, T. M.; Phung, H.-M.; Roper, T. D.; Xie, S. J. Org. Chem. 2004, 69, 3112−3215.

(28) (a) Chiacchio, U.; Saita, M. G.; Crispino, L.; Gumina, G.; Mangiafico, S.; Pistarà, V.; Romeo, G.; Piperno, A.; De Clercq, E. Tetrahedron 2006, 62, 1171−1181. (b) Chiacchio, U.; Iannazzo, D.; Piperno, A.; Romeo, R.; Romeo, G.; Rescifina, A.; Saglimbeni, M. Bioorg. Med. Chem. 2006, 14, 955−959.

(29) (a) Shin, D.; Sinkeldam, R. W.; Tor, Y. J. Am. Chem. Soc. 2011, 133, 14912−14915. (b) Hernandez, A. R.; Kool, E. T. Org. Lett. 2011, 13, 676−679. (c) Wilson, J. N.; Kool, E. T. Org. Biomol. Chem. 2006, 4, 4265−4274.

(30) Okamoto, A.; Tainaka, K.; Nishiza, K.; Saito, I. J. Am. Chem. Soc. 2005, 127, 13128−13129.

(31) Charubala, R.; Maurinsh, J.; Rösler, A.; Melguizo, M.; Jungmann, O.; Gottlieb, M.; Lehbauer, J.; Hawkins, M.; Pfleiderer, W. Nucleosides Nucleotides 1997, 16, 1369−1378.

(32) (a) Leonard, N. J.; Sprecker, M. A.; Morrice, A. G. J. Am. Chem. Soc. 1976, 98, 3987−3994. (b) Hernandez, A. R.; Kool, E. T. Org. Lett. 2011, 13, 676−679.

(33) (a) Wilhelmsson, L. M. Q. Rev. Biophys. 2010, 43, 159−183. (b) Hikida, Y.; Kimoto, M.; Yokoyama, S.; Hirao, I. Nat. Protoc. 2010, 5, 1312−1323. (c) Valis, L.; Wagenknecht, H.-A. Synlett 2005, 2281− 2284.

(34) Sekine, M.; Oeda, Y.; Iijima, Y.; Taguchi, H.; Ohkubo, A.; Seio, K. Org. Biomol. Chem. 2011, 9, 210−218.

(35) Bair, K. W.; Andrews, C. W.; Tuttle, R. L.; Knick, V. C.; Cory, M.; McKee, D. D. J. Med. Chem. 1991, 34, 1983−1990.

(36) Fukuda, S.; Kamimura, A.; Kanemasa, S.; Hori, K. Tetrahedron 2000, 56, 1637−1647.

(37) (a) Gonzalez, M. P.; Teran, C.; Fall, Y.; Teijeira, M.; Besada, P. Bioorg. Med. Chem. 2005, 13, 601−608. (b) Tchilibon, S.; Joshi, B. V.; Kim, S.-K.; Duong, H. T.; Gao, Z.-G.; Jacobson, K. A. J. Med. Chem. 2005, 48, 1745−1758. (c) Joshi, B. V.; Moon, H. R.; Fettinger, J. C.; Marquez, V. E.; Jacobson, K. A. J. Org. Chem. 2005, 70, 439−447.

(38) Collins, L. A.; Franzblau, S. G. Antimicrob. Agents Chemother. 1997, 41, 1004−1009.

(39) Grundmann, C.; Grü nanger, P. The Nitrile Oxide; Springer-Verlag: Heidelberg, Germany, 1971.

(40) North, A. C. T.; Phillips, D. C.; Mathews, F. S. Acta Crystallogr., Sect. A 1968, 24, 351−359.

(41) (a) Sheldrick, G. M. SHELXL-93: Program for Crystal Structure Analysis; University of Göttingen: Göttingen, Germany, 1993. (b) Johnson, C. K. ORTEP Report ORNL-3793; Oak Ridge National Laboratory: Oak Ridge, TN, 1965.